

# Relative Affinities of Protein–Cholesterol Interactions from Equilibrium Molecular Dynamics Simulations

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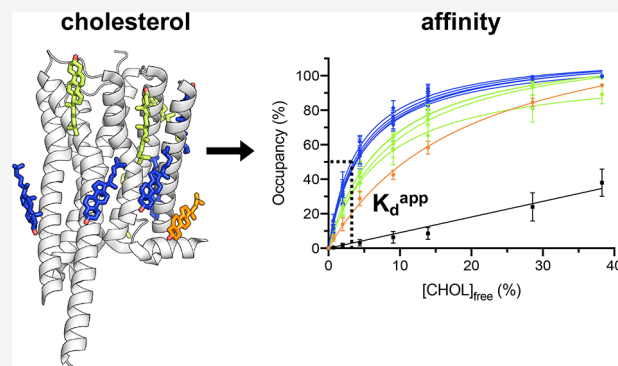


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**ABSTRACT:** Specific interactions of lipids with membrane proteins contribute to protein stability and function. Multiple lipid interactions surrounding a membrane protein are often identified in molecular dynamics (MD) simulations and are, increasingly, resolved in cryo-electron microscopy (cryo-EM) densities. Determining the relative importance of specific interaction sites is aided by determination of lipid binding affinities using experimental or simulation methods. Here, we develop a method for determining protein–lipid binding affinities from equilibrium coarse-grained MD simulations using binding saturation curves, designed to mimic experimental protocols. We apply this method to directly obtain affinities for cholesterol binding to multiple sites on a range of membrane proteins and compare our results with free energies obtained from density-based equilibrium methods and with potential of mean force calculations, getting good agreement with respect to the ranking of affinities for different sites. Thus, our binding saturation method provides a robust, high-throughput alternative for determining the relative consequence of individual sites seen in, e.g., cryo-EM derived membrane protein structures surrounded by an array of ancillary lipid densities.



## INTRODUCTION

Eukaryotic integral membrane proteins participate in a range of essential cellular functions including signaling, adhesion, solute transport, and ion homeostasis. Membrane proteins are inserted in a lipid bilayer, the composition of which varies between cellular compartments, metabolic state, and intramembrane localization.<sup>1,2</sup> Specific interactions of lipids with proteins have been observed both experimentally and in molecular dynamics (MD) simulations<sup>3–5</sup> and can alter protein functionality by, e.g., allosteric modulation<sup>6–8</sup> or bridging protein–protein oligomerization.<sup>9,10</sup>

Structural elucidation of specific protein–lipid interactions has been aided by advances in cryo-electron microscopy (cryo-EM).<sup>11,12</sup> However, distinguishing the molecular identity of lipid-like densities can be challenging, and it is limited to higher resolution examples.<sup>13,14</sup> Differentiating between phospholipid and sterol densities is somewhat easier due to their distinct shapes. In mammalian cell membranes the most abundant sterol is cholesterol, whereas in yeast and plant cell membranes it is ergosterol and phytosterol, respectively.<sup>15</sup> Cholesterol is typically present at concentrations of 30–40%,<sup>16,17</sup> although this may vary across different regions of the membrane, and is higher in sphingolipid enriched areas.<sup>18</sup> Cholesterol has been shown to bind and modulate a broad range of membrane proteins including G-protein coupled receptors (GPCRs), ion

channels, and solute transporters.<sup>6,19–23</sup> Recent cryo-EM structures have revealed several sterol-like densities surrounding protein transmembrane domains (TMDs). In these instances, the bound density is cholesterol, copurified from the native bilayer,<sup>24–26</sup> or it may correspond to cholesterol derivatives, such as cholesterol hemisuccinate (CHS), which are added during purification.<sup>27,28</sup> Often multiple cholesterol binding sites are observed within the same structure.<sup>25</sup> For example, a recent structure of the serotonin receptor, 5-HT<sub>1A</sub> (Protein Data Bank (PDB) ID 7E2X), revealed 10 cholesterol molecules surrounding the TMD, including one partially buried cholesterol adjacent to the orthosteric ligand pocket.<sup>29</sup> There is therefore a clear need to understand and characterize the relative affinities of multiple cholesterol binding sites on the same protein. However, this remains experimentally challenging, and there is a paucity of quantitative experimental biophysical data for cholesterol binding to, e.g., GPCRs<sup>30,31</sup> and other membrane proteins.

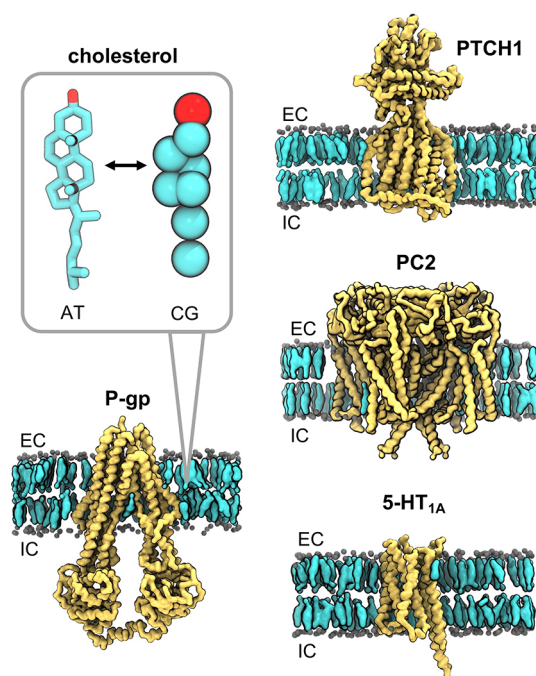
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Equilibrium MD simulations have been used extensively to expand on the information provided from structural analyses, to study protein–lipid interaction patterns, and to obtain detailed insights into specific binding sites.<sup>4,32,33</sup> In addition, biased-sampling simulations, such as potential of mean force (PMF) calculations, free energy perturbation, and metadynamics simulations, have been used to obtain lipid binding free energies, supplementing available experimental data on lipid binding affinities.<sup>34</sup> These biased simulations are often performed after initial equilibrium MD simulations, therefore requiring additional computing resources and an iterative process to select suitable reaction coordinates. This limits the applicability of such approaches to high-throughput, automated pipelines, for example, MemProtMD.<sup>35,36</sup> To circumvent these limitations, efforts have been made to derive protein–lipid binding affinities directly from equilibrium MD simulations. These have the advantage that multiple lipid sites can be simultaneously examined, such as in studies using 2D density distributions of cholesterol surrounding the  $A_2A$  and/or  $\beta_2$  adrenergic receptors, taken from either atomistic<sup>37</sup> or coarse-grained (CG) MD simulations.<sup>38</sup> Additionally, complex lipid interaction profiles<sup>32</sup> can be more readily determined, such as applied in a “density-threshold” approach with the nicotinic acetylcholine receptor in a mixed lipid environment.<sup>39</sup> This can also be achieved with biased simulations, but it requires additional simulations for each lipid species studied.<sup>40</sup> However, it remains unclear how accurate equilibrium methods are for obtaining binding affinities and whether full convergence is feasible within the limits of current MD simulations.

Here, we present a method for obtaining apparent dissociation constants ( $K_d^{\text{app}}$ 's) directly from equilibrium MD simulations. We apply this method to rank the strength of binding sites for cholesterol on three representative membrane proteins: an ATP-dependent pump (P-glycoprotein; P-gp; see below for further details), a sterol receptor/transporter protein (Patched1; PTCH1), and a member of the TRP family of ion channels (polycystin-2; PC2) (Figure 1). We investigate whether the site rankings derived from this approach are comparable with existing equilibrium and nonequilibrium methods. We also study whether these differences are maintained in the presence of higher (i.e., physiological) membrane concentrations of cholesterol. We illustrate the utility of our robust method for determining the relative affinities of multiple cholesterol sites on a membrane protein via its application to the serotonin receptor (5-HT<sub>1A</sub>), a GPCR structure recently determined by cryo-EM with 10 cholesterol molecules bound.<sup>29</sup>

## METHODS

**Equilibrium Coarse-Grained MD Simulations.** Structures of human PC2 (PDB ID 6T9N, subunits A–D),<sup>41</sup> PTCH1 (PDB ID 6RVD, subunit A),<sup>42</sup> P-gp (PDB ID 7A65, subunit A),<sup>43</sup> and 5-HT<sub>1A</sub> (PDB ID 7E2X, subunit R) were obtained from the Protein Data Bank (PDB). Nonprotein components were removed and loops were modeled using MODELER 9.20,<sup>44</sup> for Q296–N305 of PC2 (for each subunit) and L608–L732 PTCH1 (using a nine residue linker as previously described<sup>42</sup>). Proteins were converted to CG resolution using martinize.py<sup>45</sup> with an ElNeDyn 2.2<sup>46</sup> elastic network applied (spring force constant = 500 kJ mol<sup>−1</sup> nm<sup>−2</sup>, cutoff = 0.9 nm). For PC2 the elastic network was applied to each subunit separately.



**Figure 1.** Membrane proteins which bind cholesterol. Coarse-grained (CG) representations of the structures of a transporter (P-glycoprotein; P-gp; PDB ID 7A65, subunit A), a receptor (Patched1; PTCH1; PDB ID 6RVD, subunit A), an ion channel (human polycystin-2; PC2; PDB ID 6T9N, subunits A–D), and a GPCR (5-hydroxytryptamine/serotonin receptor; 5-HT<sub>1A</sub>; PDB ID 7E2X, subunit R), embedded in a phosphatidylcholine (PC; 60%) and cholesterol (40%) lipid bilayer. PC phosphate beads are shown as gray spheres, cholesterol is shown in QuickSurf representation in cyan, and proteins are in yellow. Extracellular (EC) and intracellular (IC) leaflets are labeled. The inset shows corresponding atomistic (AT) and CG representations of cholesterol with the  $\beta_3$ -hydroxyl group (equivalent to the ROH bead at CG resolution) in red.

The MARTINI2.2<sup>47</sup> force field was used to describe all components. Proteins were embedded in a symmetric 1-palmitoyl-2-oleoyl-*sn*-glycero-3-phosphocholine (POPC)/cholesterol bilayer using insane.py<sup>48</sup> (Figure 1). The following cholesterol concentrations were used with the remaining bilayer composed of POPC: 1, 2.5, 5, 10, 15, 30, and 40% cholesterol. Cholesterol was modeled using the virtual site parameters.<sup>49</sup> insane.py was also used to solvate the system with MARTINI water<sup>45</sup> before neutralization and addition of ions to ~0.15 M NaCl. Each replica was independently energy minimized using the steepest-descent method and equilibrated in 2 × 100 ns steps with restraints applied to the backbone beads.

Each protein was simulated for 5 × 5  $\mu$ s in each bilayer composition (7 bilayer compositions × 5 replicates = 175  $\mu$ s per system) using the GROMACS 2018 and 2019 simulation packages ([www.gromacs.org](http://www.gromacs.org)). A 20 fs time step was used, and periodic boundary conditions were applied. The temperature was maintained at 310 K with a V-rescale thermostat<sup>50</sup> and a  $\tau_T$  coupling constant of 1.0 ps. A Parrinello–Rahman barostat<sup>51</sup> was used to maintain the pressure at 1 bar with a  $\tau_p$  value of 12 ps and a compressibility of 3 × 10<sup>−4</sup> bar<sup>−1</sup>. Electrostatic interactions were cut off at 1.1 nm by using the reaction-field method, and Lennard-Jones interactions were cut off at 1.1 nm by using the potential-shift Verlet method. Bonds were constrained to their equilibrium values by using the LINCS algorithm.<sup>52</sup>

**Binding Site Identification.** Interactions of cholesterol with each protein were calculated using PyLipID ([github.com/wlsong/PyLipID](https://github.com/wlsong/PyLipID)).<sup>53</sup> Cholesterol interaction occupancy is defined as the fraction of simulation time where any bead of cholesterol is in contact with any bead of a protein residue, with a 0.55 nm/1.0 nm double cutoff used to define lipid contacts. PyLipID was also used to identify cholesterol binding sites by using a community analysis approach to group residues which simultaneously interact with a bound cholesterol over the course of the trajectories. This method is described in detail elsewhere<sup>54,55</sup> and has been applied to a number of recent examples to characterize lipid binding sites and kinetics.<sup>56–58</sup> Since the residue composition of sites A and B varied slightly with the percent cholesterol present in the bilayer, we selected six residues from each site, contacts to which were maintained across all cholesterol concentrations, and used these six residues in our subsequent analysis (Supplementary Figure 1). The positions of sites A and B in relation to all identified cholesterol binding sites are shown in Supplementary Figure 2 for the 40% cholesterol simulations. For 5-HT<sub>1A</sub>, sites were defined as six residues in proximity to each of the 10 modeled cholesterol densities in the structure (Supplementary Table 1).

**Binding Saturation Curves.** To define specific interactions of cholesterol with a membrane protein, we calculated the mean occupancy (eq 1) of the six selected site residues, as reported by

$$\text{occupancy} = \frac{\sum (F_x/F_t)}{n} \quad (1)$$

PyLipID,<sup>53</sup> across all cholesterol concentrations.  $F_x$  indicates the number of frames cholesterol is bound to a given residue,  $F_t$  is the total number of frames, and  $n$  indicates the total number of residues; e.g.,  $n = 6$  for interactions with a site. Nonspecific interactions were obtained by calculating the mean occupancy of residues which, in the 40% cholesterol system, had interactions within the 30–50% range. Further details regarding definitions of specific/nonspecific interactions are included in the Supporting Information and Supplementary Figure 3.

Binding saturation curves were plotted with GraphPad Prism 9.0.2 for MacOS ([www.graphpad.com](https://www.graphpad.com)). The apparent dissociation constant for cholesterol binding ( $K_d^{\text{app}}$ ) was calculated by fitting the data to eq 2, assuming site occupancies are a result of

$$\text{occupancy} = B_{\text{max}}^{\text{app}} \frac{[\text{CHOL}]_{\text{free}}}{K_d^{\text{app}} + [\text{CHOL}]_{\text{free}}} \quad (2)$$

specific interactions at one site on the protein. No constraints were used in calculation of the  $K_d^{\text{app}}$  values. Errors are reported as the standard error of the mean among five independent repeats.

The concentration of free cholesterol ( $[\text{CHOL}]_{\text{free}}$ ) was derived from the mean number of cholesterol molecules >0.8 nm from the protein surface (unbound cholesterol) as a fraction of the total number of unbound lipids (POPC and cholesterol) across simulations. Thus, our computational saturation curves circumvent approximations of free and total ligand pools often used experimentally. Note that the  $B_{\text{max}}^{\text{app}}$  values are not reported here (see the Supporting Information).

Convergence analyses were performed by re-running the fitting protocol with fewer simulations (Supplementary Figure 4) or reducing the length of the trajectory (Supplementary Figure 5).

**Density Analysis.** We adapted a previously described method used to obtain free energy values for protein–cholesterol interactions from 2D lipid density profiles observed in simulations.<sup>37,38</sup> The free energy ( $\Delta G$ ) can be then obtained by comparing the density of cholesterol bound at a specific site ( $\rho_{\text{site}}$ ) to the mean lipid density in bulk ( $\rho_{\text{bulk}}$ ) (eq 3).  $R$  denotes the gas constant in  $\text{kJ mol}^{-1} \text{K}^{-1}$  ( $8.314 \times 10^{-3}$ ) and  $T$  is the temperature in kelvin.

$$\Delta G = -RT \ln \left( \frac{\rho_{\text{site}}}{\rho_{\text{bulk}}} \right) \quad (3)$$

Our method utilizes the same underlying approach but extends the analysis to three dimensions (density in  $xyz$ ) as opposed to averaging across the bilayer normal (density in  $xy$ ). Full details of processing of the density data are provided in the Supporting Information, summarized below. Density analysis was performed with the DensityAnalysis tool implemented in MDAnalysis<sup>59,60</sup> ([www.mdanalysis.org](https://www.mdanalysis.org)) using an in-house script. Grid dimensions were fixed, and the grid center was defined as the center of mass of the protein transmembrane domain. The bin size was 0.1 nm. Three-dimensional  $\rho_{\text{site}}$  and  $\rho_{\text{bulk}}$  values were obtained by masking specific regions of the density array (Supplementary Figure 6). These values were then converted directly to free energy values by using eq 3.

**Potential of Mean Force Calculations.** The setup and analysis of PMF calculations was assisted by the pmf.py tool (DOI: <https://doi.org/10.5281/zenodo.3592318>).<sup>54</sup> CG PMF calculations were performed as described previously<sup>34</sup> in bilayers containing 30% cholesterol. Briefly, a 1D reaction coordinate was generated by pulling between the cholesterol center of mass and the backbone bead of a site residue. Windows at 0.05 nm spacing along the reaction coordinate were simulated for 1  $\mu\text{s}$  each with a  $1000 \text{ kJ mol}^{-1} \text{nm}^{-2}$  umbrella potential used to limit cholesterol movement along this coordinate. Free energy profiles were obtained by using the weighted-histogram analysis method (WHAM)<sup>61</sup> implemented in GROMACS with 2000 rounds of Bayesian bootstrapping, discarding the first 200 ns of each window. Further details are provided in the Supporting Information, and convergence of free energy values is shown in Supplementary Figure 7.

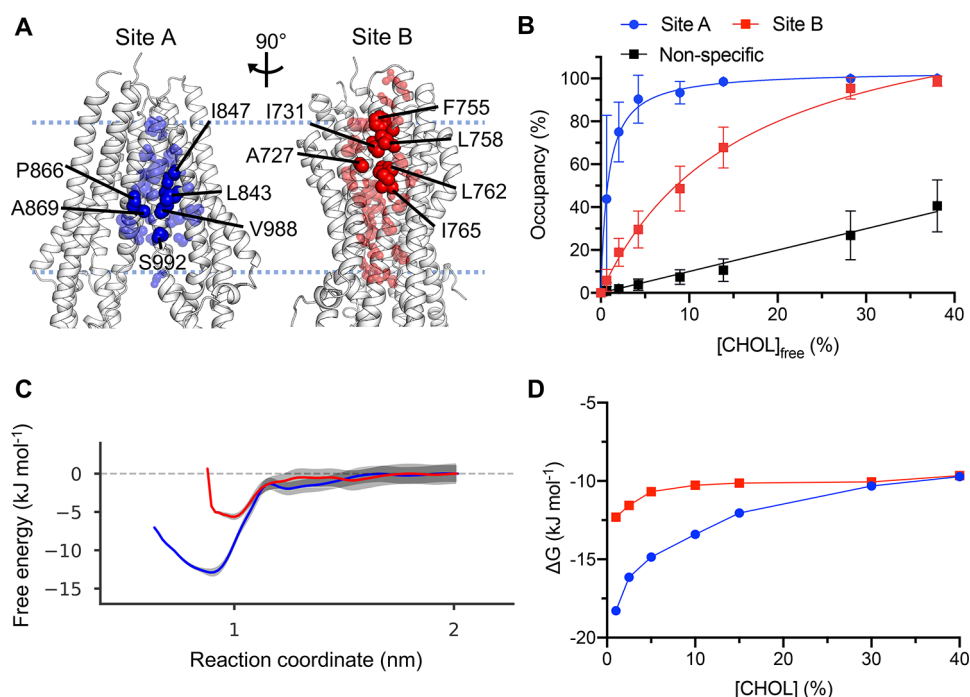
**Site Membrane Exposure.** The membrane exposure fraction was defined as the number of lipid contacts within 0.6 nm of a bound cholesterol divided by the number of total contacts (protein and lipid) to the site cholesterol, as calculated by using MDAnalysis<sup>59,60</sup> across the simulations.

## RESULTS

We set out to determine if equilibrium MD simulations are able to not only identify specific interactions of protein with lipids but also to rank the affinities of different sites, and to evaluate how well these estimates compare to biased simulations. We also wanted to assess whether values obtained from simulations were affected by the lipid concentration in the membrane.

Using equilibrium CG MD simulations, we constructed binding saturation curves, where the total cholesterol concentration was varied, and the mean occupancy of six residues in each specified binding site was determined across the concentrations of free cholesterol. The idea of this method was to mimic ligand binding assays used experimentally to produce binding saturation curves.<sup>62,63</sup>





**Figure 2.** Cholesterol binding to P-gp. (A) Cholesterol interaction sites A (blue) and B (red), identified using equilibrium simulations ( $5 \times 5 \mu\text{s}$  at each cholesterol concentration) in PC:Chol 60:40 followed by analysis using PyLipID ([github.com/wlsong/PyLipID](https://github.com/wlsong/PyLipID)).<sup>53</sup> The sites are shown mapped onto the structure of the P-gp (7A65, subunit A) TMD. Residues involved in cholesterol interactions in the 40% cholesterol simulations are shown as spheres scaled according to cholesterol residence times. The six residues selected (which were conserved across all cholesterol concentrations) are labeled (opaque), whereas the remaining residues constituting the site (in 40% cholesterol) are transparent. (B) Binding saturation curves for cholesterol binding to sites A and B across a range of cholesterol concentrations. Site occupancy was defined as the mean occupancy of the six site residues in (A). Error bars correspond to standard deviations. Nonspecific interactions were calculated from mean occupancies of specified residues with 30–50% occupancy in the 40% cholesterol simulations. (C) Free energy landscapes from potential of mean force (PMF) calculations for sites A and B from simulations in bilayers containing 30% cholesterol. Bootstrapping errors are shown in gray. (D) Free energies of binding derived from probabilities of cholesterol bound at site A (blue) or B (red) relative to the bulk probability calculated from 3D density plots of cholesterol localized surrounding P-gp.

To help with the convergence of these calculations, we chose to study the lipid cholesterol, which has been demonstrated to have relatively fast binding and/or dissociation kinetics compared to other lipids (e.g., anionic lipids such as cardiolipin and phosphatidylinositols) and hence is more amenable to sampling of multiple sites within a given simulation.<sup>33</sup> In addition, the thermodynamics of protein–cholesterol interactions have been extensively studied in both atomistic and CG simulations, with the use of both biased and unbiased methods.<sup>64</sup> These free energy estimates therefore provide a good benchmark against which to compare our results.

Three human integral membrane proteins were selected to evaluate our analysis of cholesterol interactions: the ATP-dependent efflux pump P-glycoprotein (P-gp), the proposed sterol receptor/transporter protein Patched1 (PTCH1), and the transient receptor potential (TRP) ion channel polycystin-2 (PC2) (Figure 1). In each case, cholesterol has been suggested to play a role in protein function by either allosteric modulation or direct involvement in the protein's biological process.

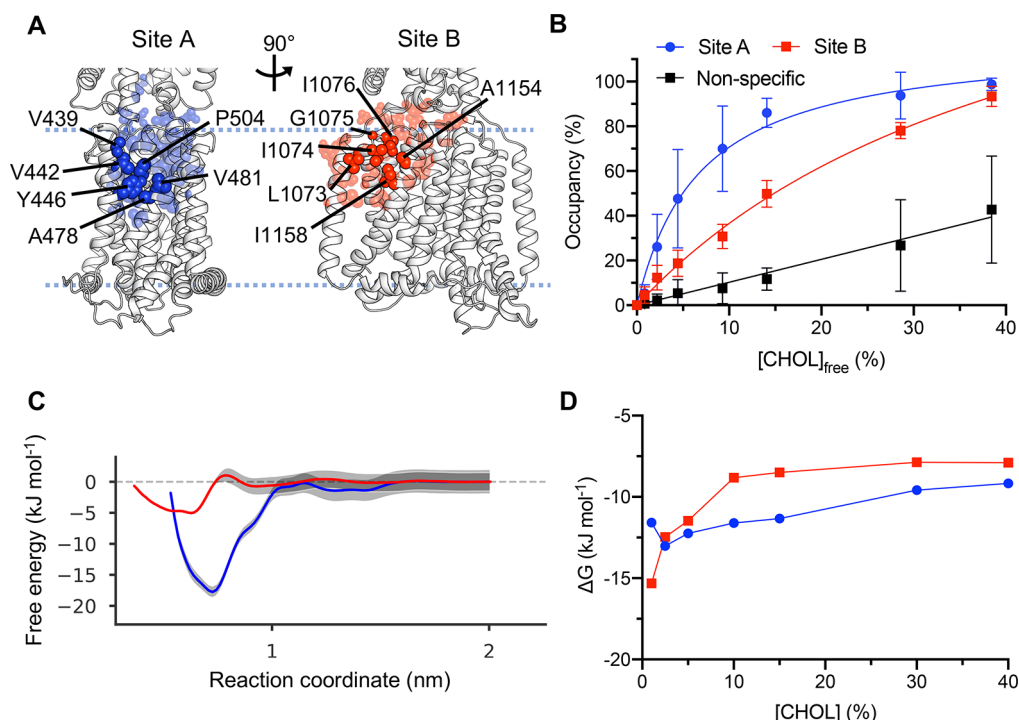
**Comparative Methods for Determining Cholesterol Binding Affinities with P-gp.** Cholesterol has been shown to alter both the drug binding properties<sup>65</sup> and ATP-mediated export rates<sup>65,66</sup> of P-gp. In addition, P-gp localizes in sphingomyelin/cholesterol enriched regions in the cell,<sup>67,68</sup> further supporting a role for cholesterol in modulating P-gp function.

Previously reported CG simulations of cholesterol binding to a human P-gp homology model observed cholesterol binding to

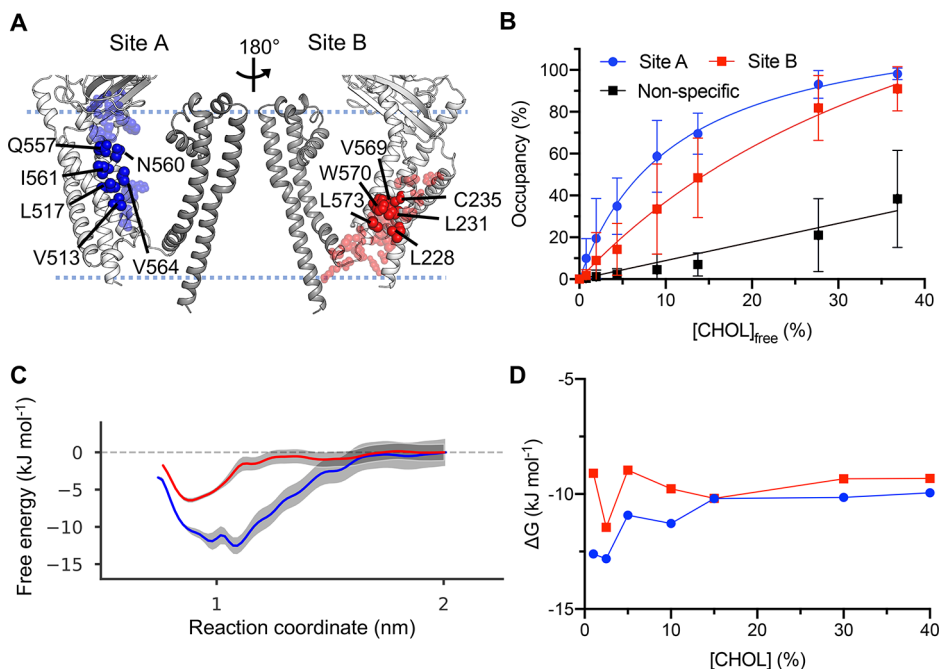
multiple sites including between TM10/TM12 and TM7/TM8, which were suggested to have different free energy values (as reported by PMF calculations).<sup>69</sup> The TM7/TM8 site was also observed in atomistic simulations of mouse P-gp.<sup>70</sup> We used PyLipID<sup>53</sup> (see Methods for details) to identify two cholesterol binding sites from equilibrium simulations. Our simulations, initiated from the recently solved human P-gp structure,<sup>43</sup> replicated the two aforementioned cholesterol binding sites from the homology model simulations. Thus, site A corresponds to cholesterol bound between TM10/TM12 and site B corresponds to cholesterol bound between TM7/TM8 (Figure 2A, Supplementary Figure 1).

Occupancies for both site A and site B increased nonlinearly with cholesterol concentration, as would be anticipated for well-defined, saturable binding sites (Figure 2B). We observe a rapid increase in site A occupancy compared to site B as cholesterol concentration is increased. The  $K_d^{\text{app}}$  of P-gp site A ( $K_d^{\text{app}} = 0.8 \pm 0.2\%$ ) is substantially lower (i.e., has a higher affinity) than that for site B ( $K_d^{\text{app}} = 16.0 \pm 0.8\%$ ). This suggests the cholesterol binding affinities of the two sites on P-gp are not equal, as is also exemplified by the variability in cholesterol affinities reported in other studies.<sup>64</sup>

We next performed PMF calculations to validate our observed differences in site affinities from our binding saturation method. For both sites we observe defined energetic wells at low reaction coordinate values, consistent with PMF profiles of other cholesterol binding sites.<sup>64</sup> We obtain free energy well depths of  $-13 \pm 2 \text{ kJ mol}^{-1}$  for cholesterol binding to site A and of  $-6 \pm$



**Figure 3.** Cholesterol binding to PTCH1. As in Figure 2 for PTCH1 (6RVD, subunit A). (A) Residues comprising cholesterol interaction sites A (blue) and B (red) on PTCH1. (B) Binding saturation curves for cholesterol binding to sites A and B as the concentration of cholesterol is varied. (C) Free energy profiles from PMF calculations for cholesterol binding to sites A and B on PTCH1. (D) Free energies derived from the probability of cholesterol binding to sites relative to in bulk, obtained using a density-based approach. For full methodological details see Figure 2.

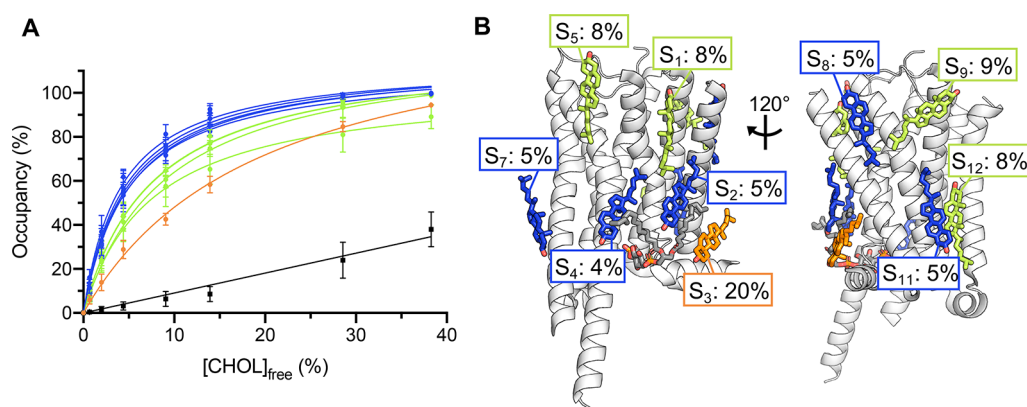


**Figure 4.** Cholesterol binding to PC2. As in Figure 2 for PC2 (6T9N, subunits A–D). For clarity, only subunit A of the PC2 homotetramer is shown in A, with the pore-lining helices of PC2 in darker gray compared to the voltage-sensing-like domain (VSLD). (A) Residues comprising cholesterol interaction sites A (blue) and B (red) on PC2. (B) Binding saturation curves for cholesterol binding to sites A and B as the concentration of cholesterol is varied. (C) Free energy profiles from PMF calculations for cholesterol binding to sites A and B on PC2. (D) Free energies derived from the probability of cholesterol binding to sites relative to in bulk, obtained using a density-based approach. For full methodological details see Figure 2.

2 kJ mol<sup>-1</sup> for site B (Figure 2C). Our PMF values are in agreement with the relative affinities of sites obtained from previous calculations on the P-gp homology model.<sup>69</sup> Thus,

both binding saturation and PMF calculations rank the sites in the same order.

We then assessed whether density-based equilibrium free energy methods could also be used to observe quantitative



**Figure 5.** Binding saturation method as applied to 10 cholesterol sites on 5-HT<sub>1A</sub>. (A) Binding saturation curves for 10 cholesterol binding sites on 5-HT<sub>1A</sub> (7E2X, subunit R). Site occupancies were obtained from the mean occupancy of six residues in proximity to the modeled cholesterol (Supplementary Table 1), as obtained using PyLipID.<sup>53</sup> Sites are colored according to the relative strength as given by the obtained  $K_d^{app}$  values (“strong”, blue; “moderate”, lime; “medium”, orange). (B) Structure of apo 5-HT<sub>1A</sub> used to obtain the binding saturation curves in (A). Cholesterol molecules are shown as sticks colored according to the relative site affinity (see (A)) and  $K_d^{app}$  values for each site indicated. Site IDs (S<sub>1</sub>–S<sub>5</sub>, S<sub>7</sub>–S<sub>9</sub>, S<sub>11</sub>–S<sub>12</sub>) correspond to those in Figure 2f of Xu et al.<sup>29</sup> The modeled phosphatidylinositol is shown in gray stick representation for reference.

differences in site binding affinities. Interestingly, for our density analysis, despite a strong difference at very low cholesterol, our free energy values for site A and site B converge at approximately  $-10 \text{ kJ mol}^{-1}$  in 40% cholesterol. This suggests some sensitivity of the method to the lipid concentration chosen for the simulation (Figure 2D).

**Extending Analysis of Cholesterol Affinities to Other Protein Examples: PTCH1 and PC2.** To test the applicability of our methods to other membrane proteins, we applied the same protocol described above in detail for P-gp to two other proteins: the receptor/transporter PTCH1 and the ion channel PC2. These are described in succession in the following sections.

**PTCH1.** Recent structural studies of PTCH1 have identified multiple sterol binding sites on the TMD and bound within the extracellular domain (ECD).<sup>71–75</sup> In addition, novel biochemical and CRISPR-based assays suggest PTCH1 alters the abundance of accessible cholesterol,<sup>74,76</sup> which collectively has led to the growing consensus that PTCH1 may function as a cholesterol transporter.<sup>77</sup>

For PTCH1, cholesterol binding sites were selected because sterol-like densities have been observed in proximity to both sites in cryo-EM structures<sup>72,75</sup> (Figure 3A, Supplementary Figure 1). Site A is localized within a structurally conserved domain formed by TM2–6 called the sterol-binding domain (SSD). Site B is situated between TM7/TM12 of PTCH1. In addition to the observed structural densities, both sites are situated at the exit points of tunnels extending through the ECD, characterized in previous atomistic simulations, and are therefore suggested to form local cholesterol binding sites for coordination of transport between the ECD and membrane.<sup>42</sup>

We again see a strong difference in the binding saturation curves for site A ( $K_d^{app} = 6.8 \pm 0.3\%$ ) and site B ( $K_d^{app} = 46 \pm 2\%$ ) (Figure 3B), suggesting that site A has a far higher apparent affinity than site B, although the difference between sites was somewhat less than seen in P-gp.

This difference is also reflected in our PMF calculations, from which we obtain free energy well depths of  $-18 \pm 3 \text{ kJ mol}^{-1}$  for cholesterol binding to site A and  $-6 \pm 1 \text{ kJ mol}^{-1}$  for site B (Figure 3C). This suggests that we can obtain qualitative agreement between the ranking of site affinities using binding saturation curves and density analysis compared to PMFs and

that the magnitudes can be compared between proteins and appear to reflect genuine differences in site affinities.

These differences are reflected in our 3D density analysis, albeit with a muted difference between the sites, which at  $>15\%$  cholesterol gives values of about  $-10 \text{ kJ mol}^{-1}$  for site A and  $-8 \text{ kJ mol}^{-1}$  (40% cholesterol) for site B (Figure 3D).

**PC2.** A combined cryo-EM and MD study of PC2 identified cholesterol-like density located between the voltage-sensing-like domain (VSLD) and the pore helices, which coincided with a cholesterol binding site seen in CG simulations.<sup>41</sup> In addition, both PC2 and PTCH1 localize within the primary cilia of cells where levels of accessible cholesterol are regulated and where cholesterol has been shown to play roles in initiating intracellular signaling pathways.<sup>78</sup>

As before, we identified cholesterol binding sites on PC2 and constructed binding saturation curves. Site A on TM3/TM4 (Figure 4A, Supplementary Figure 1) has previously been identified from a combined structural and simulation study.<sup>41</sup> Site B is on the interface of TM1/TM4. We again see differences in site affinity between site A ( $K_d^{app} = 11 \pm 1\%$ ) and site B ( $K_d^{app} = 49 \pm 9\%$ ) from our saturation curves (Figure 4B). The  $K_d^{app}$  for site A is higher than that observed for P-gp or PTCH1.

From PMF calculations we obtain a free energy well depth of  $-13 \pm 3 \text{ kJ mol}^{-1}$  for cholesterol binding to site A, in agreement with a previously reported value of  $-12 \pm 3 \text{ kJ mol}^{-1}$  for cholesterol binding to this site obtained by a similar method<sup>41</sup> (Figure 4C). For site B we obtain a free energy value of  $-7 \pm 1 \text{ kJ mol}^{-1}$ , consistent with differences in site affinities from the saturation curves.

Finally, from the density analysis we observe stabilization of the free energy values at  $>15\%$  cholesterol, corresponding to approximately  $-10$  and  $-9 \text{ kJ mol}^{-1}$  for sites A and B, respectively (Figure 4D).

#### Affinities of Multiple Cholesterol Sites on One Protein.

We sought to assess whether the binding saturation method could be applied to a membrane protein with several bound cholesterol molecules, exploiting the method's ability to obtain multiple  $K_d^{app}$ 's from the same simulation data set. For this we chose a recent structure of the 5-HT<sub>1A</sub> GPCR, determined in complex with 10 cholesterol molecules.<sup>29</sup> Here, we used the structurally observed cholesterol densities to define the position of the binding sites (site IDs as numbered in ref 29,



Supplementary Table 1) and constructed binding saturation curves for each site.

We observe saturable binding curves for all 10 sites, validating the position of modeled cholesterol densities in the 5-HT<sub>1A</sub> structure<sup>29</sup> (Figure 5A). All except one site (S<sub>3</sub>) had  $K_d^{app}$ 's ranging between 4 and 9%, similar to site A  $K_d^{app}$ 's for P-gp, PTCH1, and PC2. These binding sites could be further separated into two subcategories: “strong” sites with  $K_d^{app}$ 's of 4–5% (S<sub>2</sub>, S<sub>4</sub>, S<sub>7</sub>, S<sub>8</sub>, S<sub>11</sub>) (Figure 5B, blue; Supplementary Table 1) and “moderate” sites with  $K_d^{app}$ 's of 8–9% (S<sub>1</sub>, S<sub>5</sub>, S<sub>9</sub>, S<sub>12</sub>) (Figure 5B, lime) which had distinct binding saturation profiles (Figure 5A). The remaining site (S<sub>3</sub>) (Figure 5B, orange) had an intermediate affinity ( $K_d^{app}$  = 20%) relative to sites A and B of P-gp, PTCH1, and PC2, suggestive of a “medium” affinity site. Thus, using a single set of simulations, we are able to rank the respective affinities of the 10 cholesterol molecules bound to 5-HT<sub>1A</sub>.

## DISCUSSION

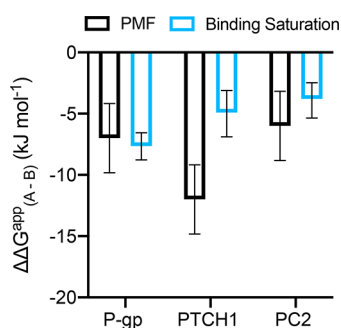
Increasingly, structures and simulations reveal a range of lipids bound to sites on the TMDs of membrane proteins.<sup>25,29,32</sup> Nevertheless, challenges with structural interpretation prevail when attempting to assign meaning to bound lipids in a biological context and/or for protein function. Ranking the affinity of lipid sites can aid this interpretation by establishing which sites may be more relevant/prevalent in a biological context.

We compared the affinities of two cholesterol sites on each of P-gp, PTCH1, and PC2 using equilibrium and biased MD simulations. Calculating the difference in apparent site free energies ( $\Delta\Delta G^{app}$ ) between sites A and B (eq 4) reveals good

$$\Delta\Delta G^{app} = \Delta G_{site A}^{app} - \Delta G_{site B}^{app};$$

$$\Delta G_{site}^{app} = -RT \ln(K_d^{app}) \quad (4)$$

agreement between the ranking of site affinities derived from PMF calculations and from our binding saturation method (Figure 6), i.e.,  $\Delta\Delta G^{app} < 0$  for both methods. Furthermore, the magnitude of  $\Delta\Delta G^{app}$  between methods agrees well for P-gp and PC2, which yield  $\Delta\Delta G^{app}$  values of  $-4$  to  $-8$  kJ mol<sup>-1</sup>.  $\Delta\Delta G^{app}$  values for PTCH1 differ somewhat between methods, which we attribute to a site A free energy much greater than observed for



**Figure 6.** Difference in apparent free energies of binding, site A – site B. For each protein,  $\Delta\Delta G^{app}$  (site A – site B) is shown, estimated from the difference in PMF well depth (black; Figures 2C, 3C, and 4C) and from the difference in  $\Delta G^{app} = -RT \ln K_d^{app}$  (light blue; using  $K_d^{app}$  values obtained from fitting the binding saturation curves in Figures 2B, 3B, and 4B). PMF errors were calculated in quadrature (total error =  $\sqrt{A_{err}^2 + B_{err}^2}$ ) since PMFs for sites A and B are independent.

cholesterol binding to other proteins by PMF calculations.<sup>64</sup> Thus, this suggests our binding saturation method can accurately rank the order and magnitude of site affinities when compared to robust free energy methods.

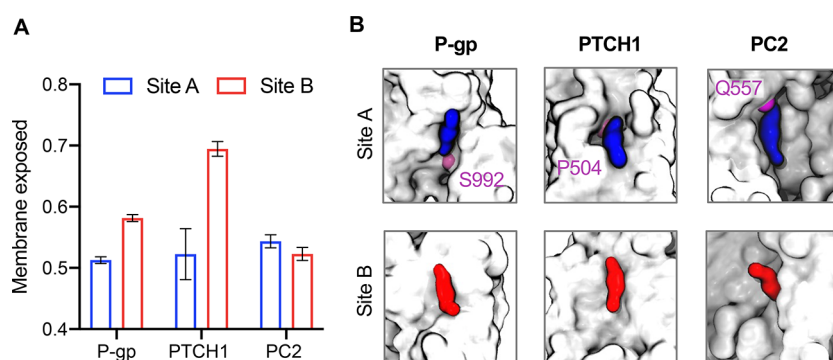
It is worth noting is that we do not see strict agreement between all obtained  $K_d^{app}$  and PMF values. One notable example of this is the P-gp site A, which gives the strongest  $K_d^{app}$  but not the highest PMF value. Interestingly, previous PMFs for this system have been much higher,<sup>69</sup> and more in line with our  $K_d^{app}$ . This points to a variance in the PMF calculations depending on the conditions used, something we do not expect to factor into our  $K_d^{app}$  calculations.

The *in silico* binding saturation method circumvents two approximations that are routinely applied in equivalent experimental procedures. First, our cholesterol binding occupancies are specific to the site of interest, avoiding complications created by conflating micro and macro dissociation constants, and second, the concentration of free cholesterol can be directly calculated rather than approximating to the total cholesterol concentration. This allows for differences in site affinities to be observed over a range of physiologically relevant free cholesterol concentrations. We note that care should be taken when determining site affinities from density-based equilibrium methods as these appeared to show some sensitivity to the overall lipid concentration in the membrane (Figures 2D, 3D, and 4D) and/or the degree of site sampling at low percent cholesterol.

We note that, for PTCH1 and P-gp, the free energy values obtained from PMFs were broadly similar at different membrane cholesterol concentrations (Supplementary Figure 8). This suggests that the site A free energy value was not affected by whether site B was occupied (as in 30% cholesterol, Figures 2C, 3C, and 4C) or unoccupied (in 0% cholesterol, Supplementary Figure 8). Conversely, the PC2 site PMF was considerably higher at 30% cholesterol than at 0% cholesterol. This is potentially due to allosteric coupling between cholesterol sites. However, it is unclear how well this could be modeled in CG simulations with an elastic network model. Alternatively, there may be direct cooperativity between cholesterol molecules, with overlapping cholesterol sites. Either way, this makes interpretation of any value taken from free energy calculations difficult, as the presence or absence of other cholesterol could impact the finding. The presented binding saturation method, however, implicitly takes this into consideration.

The computational cost and ease of setup are key considerations if we wish to investigate affinities using high-throughput simulation methods applied to a wide range of membrane proteins. For the PMFs, each site on a given protein was simulated for approximately  $65 \times 1 \mu s$  umbrella sampling windows, in addition to the initial steered MD simulations ( $\sim 0.03 \mu s$ ), from which those windows were derived. Thus, approximately  $130 \mu s$  of CG simulation time was used in the PMF calculations to derive the two site affinities on a protein, at a single cholesterol concentration.

For the binding saturation method, the total CG simulation time for both sites was  $175 \mu s$  ( $5 \times 5 \mu s$  at 7 lipid concentrations) across all free cholesterol concentrations. The binding saturation method was surprisingly robust, with as few as two replicas required to reach  $K_d^{app}$  convergence and one replica sufficient to observe qualitative differences in site affinities (Supplementary Figure 4). In addition,  $3 \mu s$  per simulation appears to be sufficient for convergence (Supplementary Figure 5). Therefore, the total simulation time could be reduced to  $21\text{--}42 \mu s$  and still



**Figure 7.** Molecular basis of observed differences in site affinities. (A) Fractional membrane exposure of the bound site A and site B cholesterol for P-gp, PTCH1, and PC2 across simulations. Membrane exposure was defined as the number of lipid contacts within 0.6 nm of the bound cholesterol divided by the total number of contacts (protein and lipid) within 0.6 nm. Error bars indicate the standard error of the mean across replicates. (B) Binding pose of cholesterol bound to sites A (blue) and B (red) of P-gp, PTCH1, and PC2, as obtained using PyLipID<sup>53</sup> from our CG simulation data. Site cholesterol is shown bound to the surface (all beads, white) of the proteins, indicating differential burial of the site cholesterol. The locations of polar residues in proximity to the ROH bead of site A cholesterol are indicated in purple.

yield quantitative differences in site affinities. Furthermore, the number of residues used to define the high affinity site (site A) could be reduced from six to one, reducing the amount of user input required in simulation analysis (Supplementary Figure 9). The lower affinity site (site B) was more sensitive to the number of site residues, as expected for weaker site binding. Setting up equilibrium simulations is more amenable to automation compared to the careful selection of reaction coordinates required in biased methods, making the former approach suitable for use in high-throughput pipelines. Crucially, equilibrium methods allow multiple site affinities to be obtained from the same simulation data set, meaning that analyses could be extended to many sites in the same system for the same computational cost. We exemplify this here, to obtain relative affinities of 10 cholesterol sites on the 5-HT<sub>1A</sub> receptor. From binding saturation curves, we can group these sites into three categories corresponding to “strong”, “moderate”, and “medium” affinity sites compared to  $K_d^{app}$ s of sites on P-gp, PTCH1, and PC2 (Figure 5). Performing 10 equivalent PMF calculations would require  $\sim 650 \mu s$  of simulation time,  $\sim 4$  times the equilibrium CG simulation time used here. Thus, the binding saturation method is a suitable alternative for investigating site affinities, yielding tractable and accurate results with modest input required from the user.

**What Dictates Differences in Cholesterol Binding Affinities?** We sought to understand whether key structural features between site A and site B underpin the observed differences in site affinities (Figure 7). For P-gp and PTCH1, the membrane exposure of the bound cholesterol was lower for site A than for site B (Figure 7A). This suggests that the more buried the cholesterol site is, the higher the observed affinity. That said, the degree of site exposure to the surrounding membrane was not sufficient to fully describe differences in site affinity for PC2, where both sites were similarly buried but the affinities were different. For PC2, the presence of a polar residue (Q557) in proximity to the hydroxyl (ROH) bead of cholesterol appears to enhance the affinity of cholesterol binding to site A (Figure 7B). Equivalent polar residues are not present in site B (Figure 4A). A polar residue was also present in site A of P-gp and that of PTCH1 adjacent to the cholesterol ROH bead (Figure 7B). Thus, “strong” cholesterol binding is enhanced by both the pocket-like nature of a binding cavity and polar residues in direct contact with the lipid headgroup.

For 5-HT<sub>1A</sub>, almost all sites contained polar residues in proximity to the cholesterol hydroxyl, reflected in the low  $K_d^{app}$  values obtained from the binding saturation curves (Figure 5, Supplementary Table 1). Higher affinity sites are more likely to persist during the relatively harsh purification and solubilization process used to obtain membrane protein structures by cryo-EM, consistent with the high affinity of sites observed on 5-HT<sub>1A</sub>. One site on 5-HT<sub>1A</sub> (S<sub>1</sub>) is proposed to stabilize the orthosteric ligand binding pocket and regulate binding of aripiprazole to the receptor.<sup>29</sup> We obtained a  $K_d^{app}$  value of 8% for this site which, while possessing a reasonably high affinity, was not the strongest cholesterol binding site on 5-HT<sub>1A</sub>. Reduced affinity at S<sub>1</sub> may assist dynamic binding/unbinding of cholesterol to this site compared to a constitutively occupied, higher affinity, binding site.

Perhaps a more intriguing question is why, from a functional perspective, membrane proteins might show differences in cholesterol affinities across their surfaces. Differential site affinities on proteins could be utilized, e.g., for cholesterol-dependent differences in protein regulation. PC2 and PTCH1 localize to the primary cilia, where the abundance of accessible membrane cholesterol is highly regulated.<sup>79</sup> Changes in cilia cholesterol levels coincide with activation levels of key signaling pathways and to the subcellular localization of PTCH1.<sup>76,80</sup> In addition, the abundance of membrane cholesterol within organelles increases between the endoplasmic reticulum and the plasma membrane.<sup>17</sup> Cholesterol binding/unbinding to sites could therefore aid in protein trafficking to its native membrane environment. For example, the dynamic localization of SNARE proteins within the trans-Golgi network and endosomes is affected by membrane cholesterol abundance, affecting SNARE recycling between membranes.<sup>81</sup>

One factor not considered in the study is the ability of other specific lipids to influence the affinity of a different lipid to a site. For example, the presence of PIP<sub>2</sub> in a complex membrane environment enhances the affinity of PS binding to the Kir2.2 channel.<sup>54</sup> Additionally, we do not assess the relative affinity of different lipids binding to the same site as has been investigated in a recent study of the Kir6.2 channel.<sup>40</sup> Future work will seek to evaluate how changes in cholesterol concentrations influence site affinities within the context of more realistic membrane environments and to assess roles lipid synergy might play in affinity modulation.



In summary, we have evaluated the binding affinities of cholesterol to two sites on a range of proteins, drawing comparisons between well-established PMF and equilibrium methods. We describe a novel binding saturation curve method for obtaining affinities from equilibrium simulations, intended to imitate experimental binding assays. This method was also applied to simultaneously probe the affinities of 10 cholesterol binding sites on a protein, demonstrating how the method could be scaled for automated and/or high-throughput analysis. The binding saturation method accurately ranks the order and relative magnitude of site affinities when compared to PMF calculations and could be readily applied to study affinities of other lipid/ligand binding events.

## ■ ASSOCIATED CONTENT

### SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.jctc.1c00547>.

Detailed descriptions of how the binding saturation, PMF, and density analyses are set up and analyzed, as well as the full data to complement Figure 5 (PDF)

Views of cholesterol bound to the sites analyzed; all cholesterol sites; definitions of specific and nonspecific sites; convergence of replicates; convergence with time; 3D density analysis; PMF convergence; PMFs with different cholesterol percentages present; binding site sensitivity to number of residues per site (PDF)

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## Notes

The authors declare no competing financial interest.

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